

Figure S1. *late flowering in pGAS1::FT ft-10 tsf-1 (lgf)* mutants and characterization of *lgf58*. (A) *lgf* mutants identified in a sensitized genetic screen. Bar: 1 cm. (B) Comparison of flowering time of identified *lgf* mutants. (C) Phenotype of *lgf58* compared to pGAS1::FT *ft-10 tsf-1*, Col-0 and *pny-40126* mutant. *lgf58* showed short stature, lanceolated leaves, phyllotactic abnormalities and late flowering. (D) Quantification by RT-qPCR of expression levels of FT-transcriptionally regulated genes in *lgf58* mutant. Error bars in (B) and (C) indicate s.d. Letters shared in common between the genotypes indicate no significant difference (t-test, $P < 0.05$). Asterisks indicate statistical differences between Col-0 and other genotypes (t-test; $P < 0.05$)

A*lgf58* x pGAS1::FT *ft-10 tsf-1*

BC1F1



566 BC1F2



174 late flowering



Pool of gDNA



Illumina sequencing



SHOREmap backcross analysis

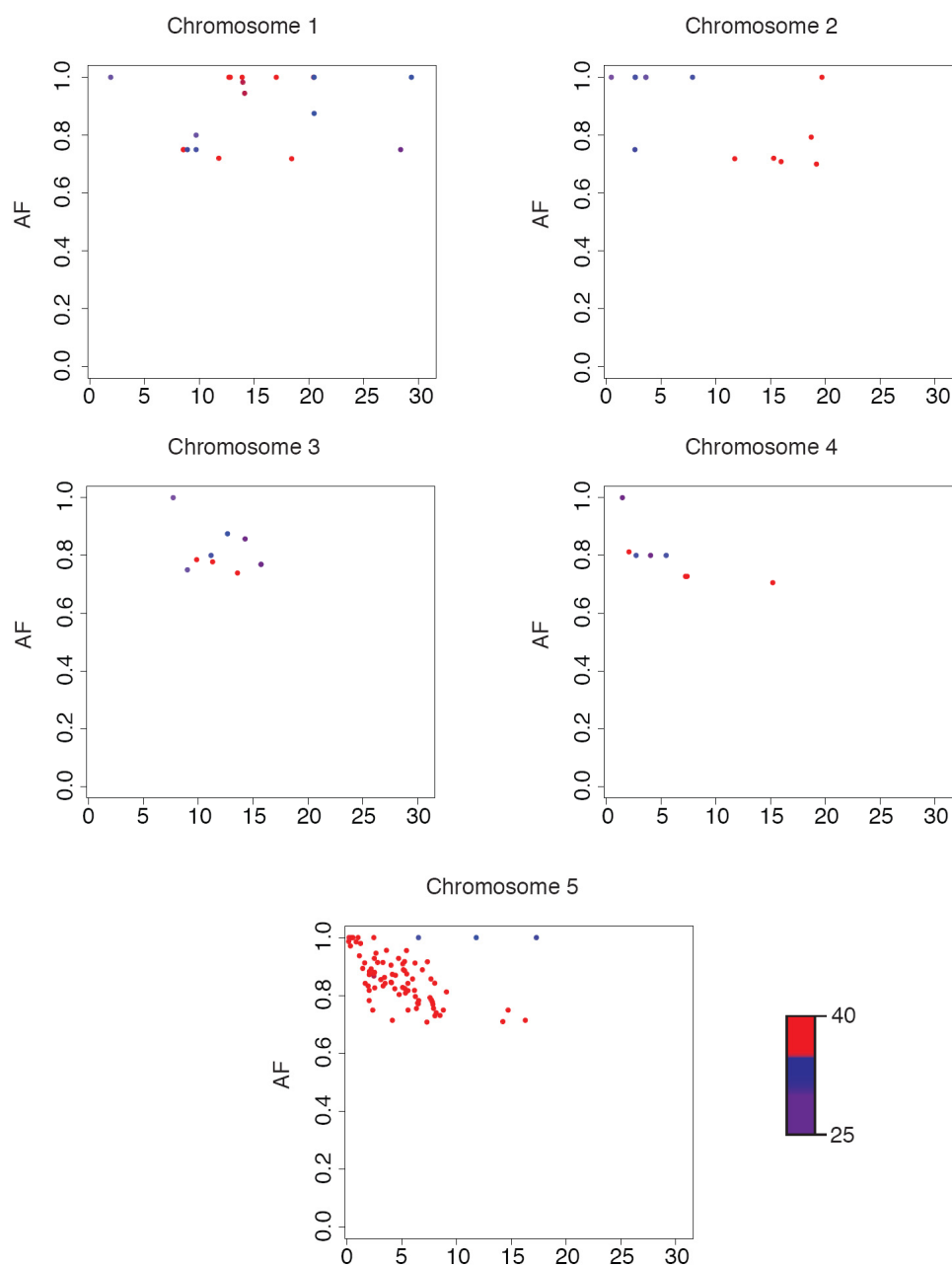
B

Figure S2. Cloning-by-sequencing of *lgf58* mutation. (A) Workflow employed to identify the causal mutation in the *lgf58* mutant. (B) Graphics showing the allelic frequency estimations at EMS-induced mutations (AF, y axis) across the five chromosomes (Mb, x axis) of *lgf58*. AFs were calculated dividing the number of reads supporting the mutant allele by the number of all reads aligning to a given marker. The color code indicates the resequencing consensus (SHORE) score. EMS-mutations showing a SHORE score higher than 25 were selected. AFs in chromosome 5 were higher as compared with other regions in the genome.

A

Col-0

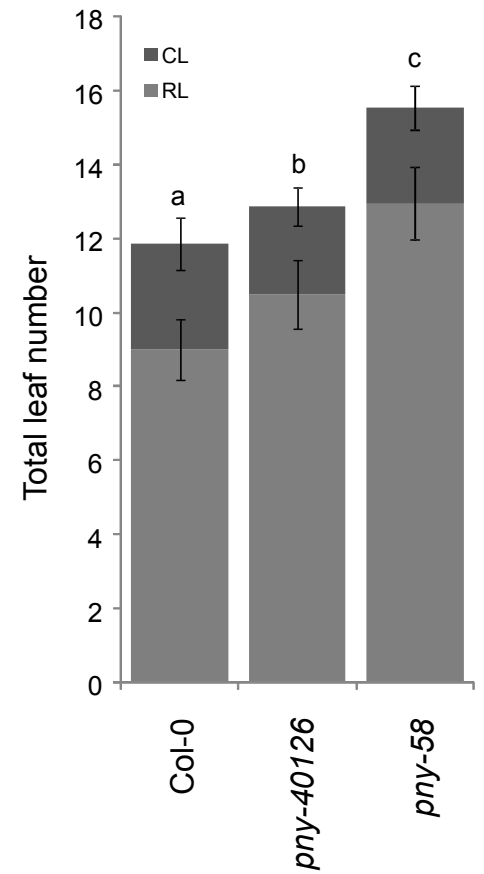
*pny-58***B**

Figure S3. Characterization of mutant plants carrying the *pny-58* allele. (A) Phenotypic comparison between Col-0 and *pny-58*. *pny-58* displayed late flowering, short stature and phyllotactic abnormalities (B) Comparison of flowering time between *pny* mutant plants. *pny-40126* (Smith et al., 2003) and *pny-58* mutant plants. Error bars indicate s.d. Letters shared in common between the genotypes indicate no significant difference (t-test, $P < 0.05$).

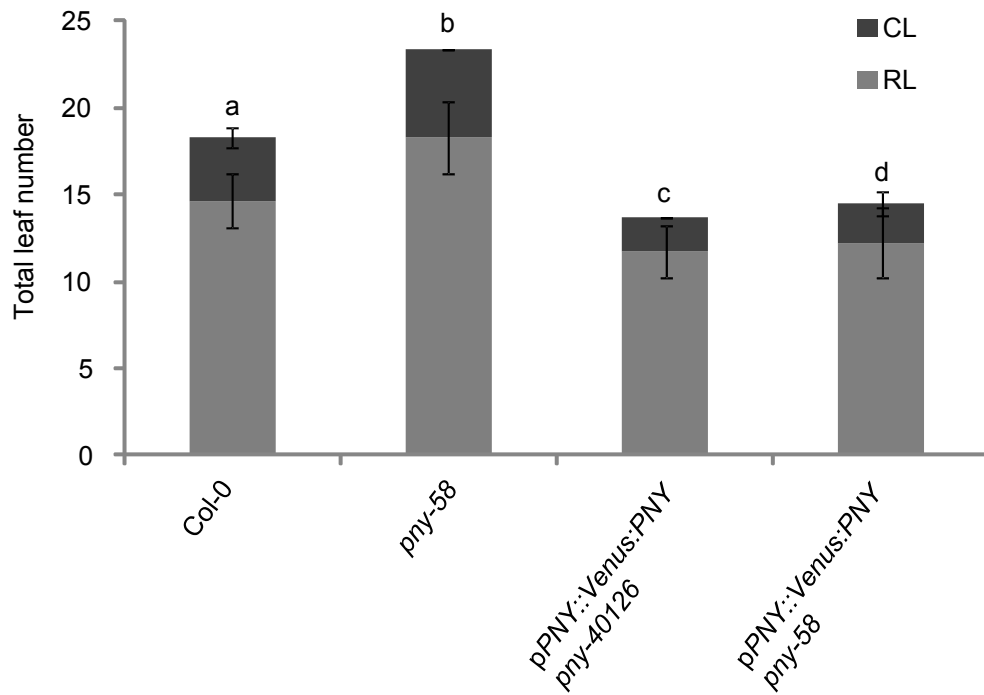
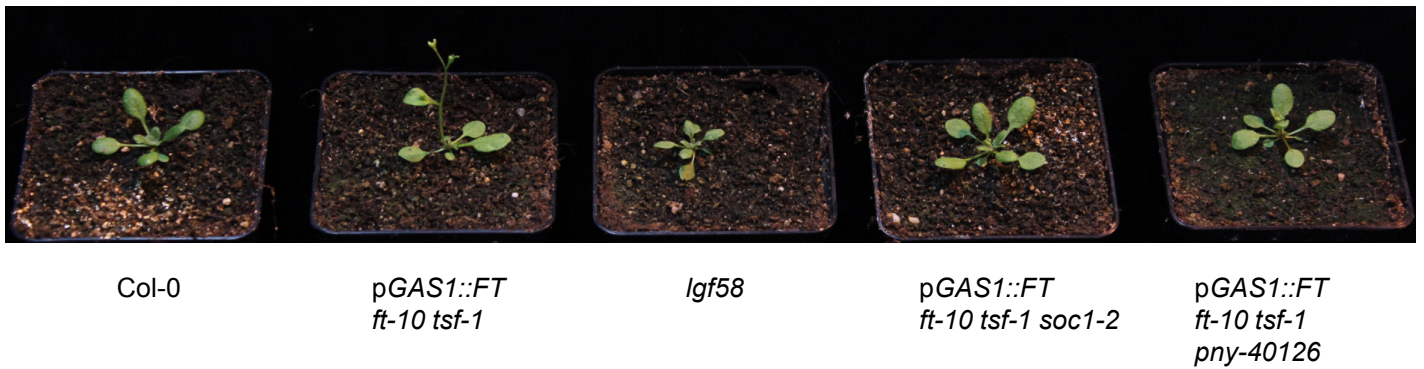
A**B**

Figure S4. *PNY* mutations caused suppression of FT. (A) Complementation assay of *pny-58* mutant. *pPNY::Venus:PNY* was crossed to *pny-58*. F2 *pny-58* plants carrying the transgene recapitulated early flowering. (B) The *PNY* mutant allele *pny-40126* in *pGAS1::FT ft-10 tsf-1* caused late flowering. Error bars indicate s.d. Letters shared in common between the genotypes indicate no significant difference (t-test, $P < 0.05$).

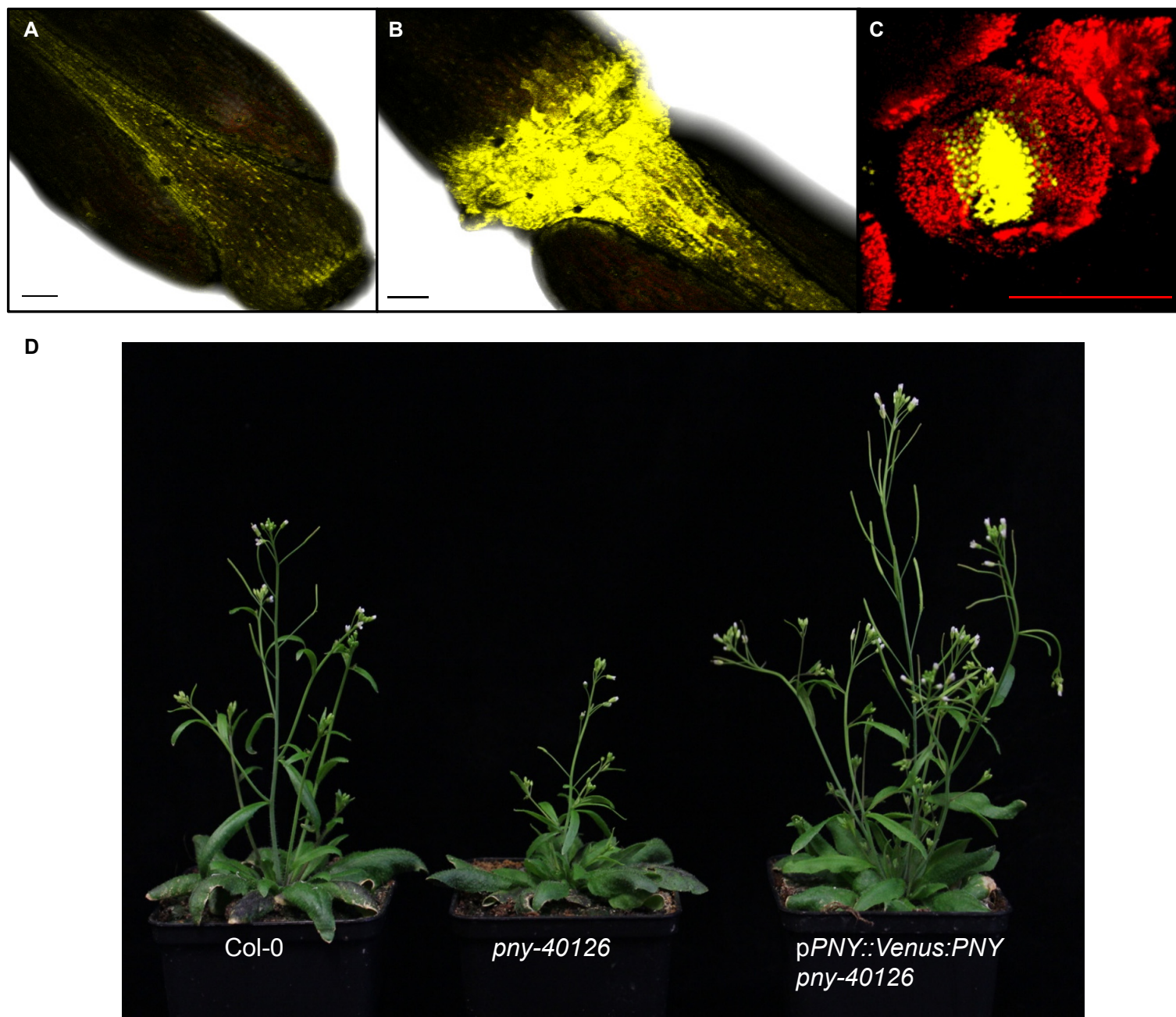


Figure S5. Functional characterization of pPNY::Venus:PNY. Expression of pPNY::Venus:PNY in the distal (A) and proximal (B) fruit regions and the central region of a young floral bud (C). Complementation assay of *pny-40126* mutant with pPNY::Venus:PNY. pPNY::Venus:PNY restored the wild type phenotype in the *pny-40126* mutant. Scale bars: 100 μ M.

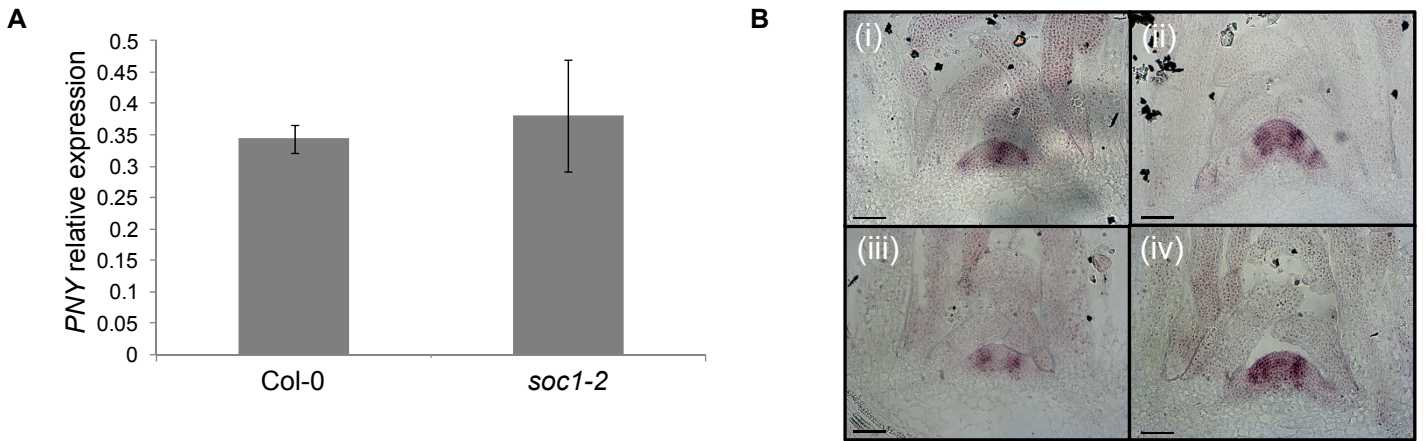


Figure S6. Photoperiod control of *PNY* pattern of expression. (A) Expression levels of *PNY* in *soc1-2* mutant compared to Col-0. Plants were grown under SDs for two weeks. Aerial parts were used for RNA extraction. Error bars indicate s.d. (B) Pattern of expression of *PNY* in Col-0 (i and ii) and *ft-10 tsf-1* mutants (iii and iv) at vegetative (i and iii) and reproductive stages (ii and iv). Scale bars: 50 μ M.

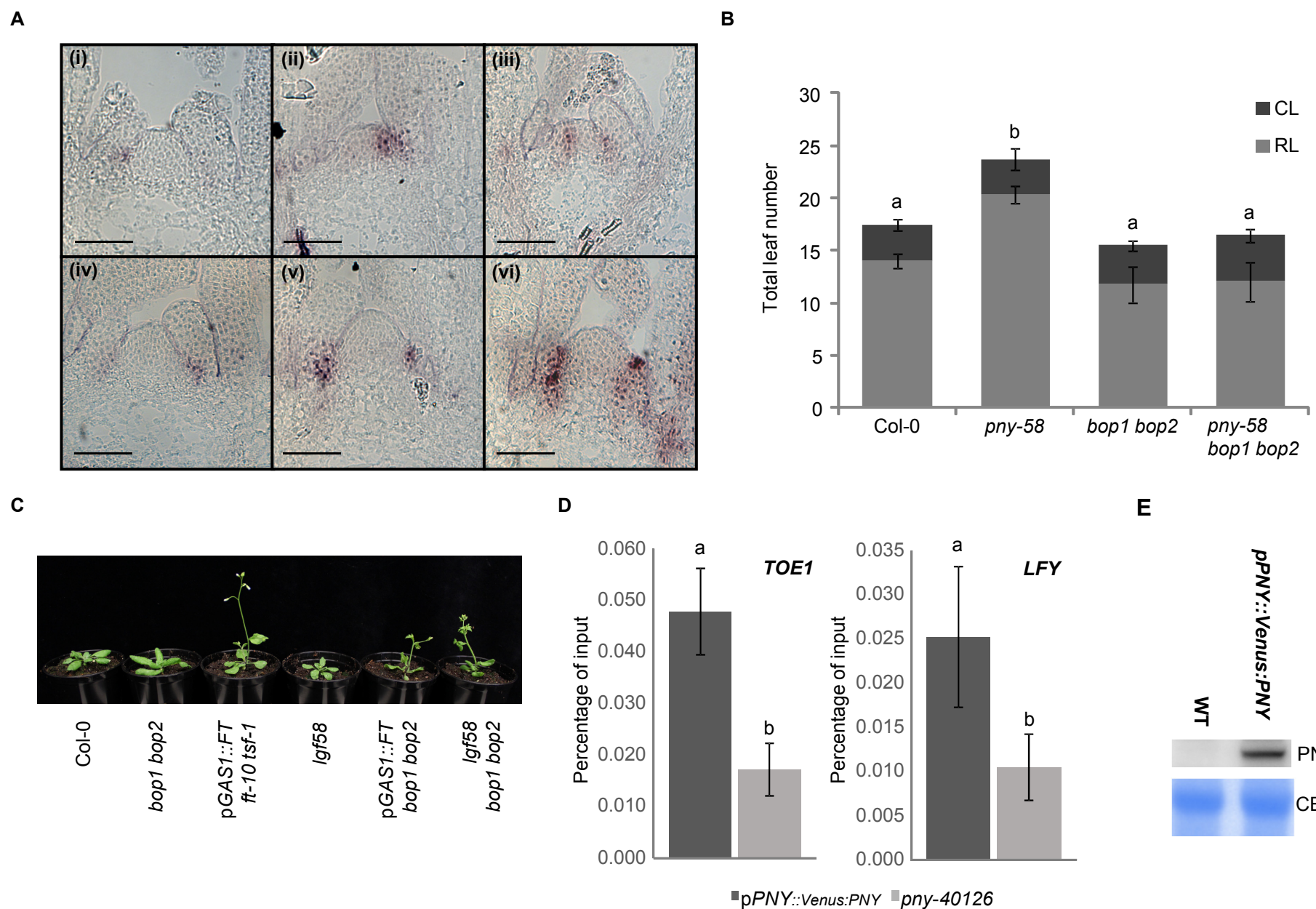
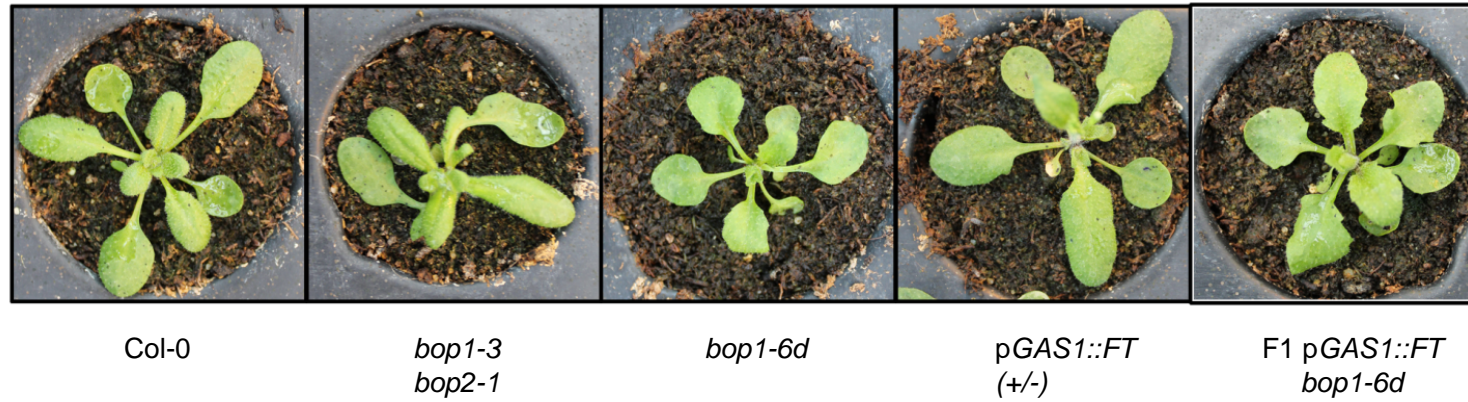
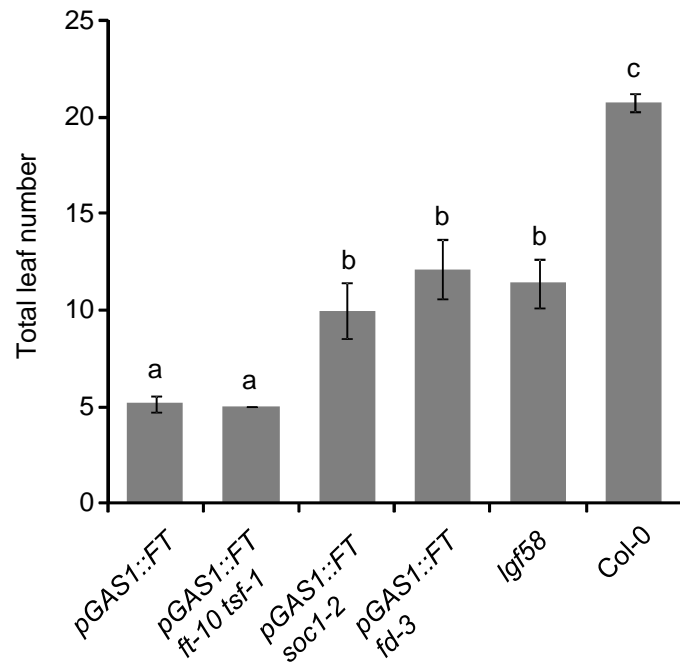


Figure S7. *PNY* controls expression of *BOP1/2* genes. (A) Expression pattern of *BOP2* in shoot meristems of Col-0 (i and iv), *pny-58* (ii and v) and *pny-40126* (iii and vi) during vegetative (I, ii and iii) and floral transition (iv, v and vi) stages. Scale bars: 50 μ m. (B) Flowering time of the triple mutants *pny-58 bop1-3 bop2-1*. Letters shared in common between the genotypes indicate no significant difference in flowering time (ANOVA test, Holm-Sidak method, $P = 0.05$). (C) Picture of plants carrying various mutant combinations for *PNY*, *BOP1/2* and *FT*. (D) ChIP-qPCR to test *PNY* binding on *TOE1* and *LFY*. Error bars in (B) and (D) indicate s.d. Letters shared in common between the genotypes indicate no significant difference (t-test, $P < 0.05$). (E) Venus:PNY protein levels were detected by immunoblotting assay using an anti-GFP antibody (upper panel) in WT and pPNY:: Venus:PNY inflorescences. CBB, Coomassie Brilliant Blue was used as a loading control (lower panel).

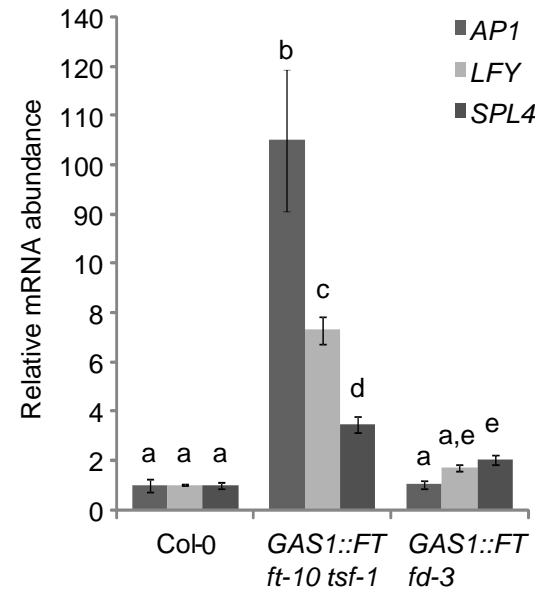
A



B



C



D

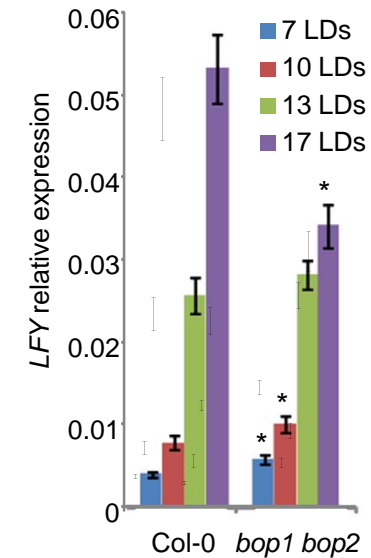


Figure S8. *BOP1/2* genes interfere *FT* signaling pathway by affecting *FD* expression. (A) Phenotypes of plants misexpressing *BOP* genes. (B) Flowering time of different suppressors *GAS1::FT* (C) Expression levels of *FT*-transcriptional regulated genes in *GAS1::FT fd-3* mutant. Letters shared in common between the genotypes indicate no significant difference (t-test, $P < 0.05$). Expression levels of *LFY* in wild type and *bop1 bop2* plants at different shoot meristem developmental stages. RNA was extracted from shoot apices of plants grown during 7 LDs (vegetative stage), 10-13 LDs (floral transition) and 17 LDs (reproductive stage). Asterisks indicate statistical differences between *Col-0* and *bop1 bop2* (t-test, $P < 0.05$). Error bars in (B), (C) and (D) indicate s.d.

Supplemental Table I. Candidate loci identified by SHOREmap

Chr ⁽¹⁾	Pos ⁽²⁾	R ⁽³⁾	M ⁽⁴⁾	N ⁽⁵⁾	AF ⁽⁶⁾	Sh ⁽⁷⁾	Region ⁽⁸⁾	Gene ID ⁽⁹⁾	Type ⁽¹⁰⁾	R ⁽¹¹⁾	M ⁽¹²⁾
5	149076	C	T	74	0.99	40	intronic/n oncoding	AT5G01360			
5	185516	C	T	39	1	40	intronic/n oncoding	AT5G01450			
5	318400	C	T	34	0.97	40	intergenic				
5	355398	C	T	5	1	32	intergenic intronic/n				
5	369679	C	T	73	1	40	oncoding	AT5G01950			
5	397204	C	T	61	1	40	CDS	AT5G02030	Nonsyn	Q	STOP
5	407433	C	T	78	1	40	CDS	AT5G02070	Nonsyn	G	R
5	458448	C	T	91	1	40	CDS	AT5G02250	Nonsyn	L	F
5	543204	C	T	51	1	40	intronic/n oncoding	AT5G02470			
5	832812	C	T	69	0.99	40	CDS	AT5G03380	Nonsyn	G	E
5	1003048	C	T	23	1	40	intergenic				
5	1130080	C	T	60	0.94	40	CDS	AT5G04140	Nonsyn	P	L
5	1228282	C	T	49	0.98	40	intronic/n oncoding	AT5G04360			
5	1432099	C	T	42	0.89	40	intronic/n oncoding	AT5G04895			
5	1610524	C	T	21	0.91	40	intergenic				
5	1656838	C	T	32	0.84	40	CDS	AT5G05570	Nonsyn	R	W
5	1934078	C	T	25	0.83	40	intergenic				
5	2016343	C	T	36	0.78	40	CDS	AT5G06580	Nonsyn	R	K
5	2017876	C	T	45	0.82	40	CDS	AT5G06590	Nonsyn	P	S
5	2027212	C	T	48	0.87	40	CDS	AT5G06600	Nonsyn	A	T
5	2051145	C	T	53	0.88	40	CDS	AT5G06670	Syn	L	L
5	2197116	C	T	58	0.89	40	CDS	AT5G07070	Nonsyn	G	E
5	2355749	C	T	9	0.75	40	intergenic				
5	2444272	C	T	17	1	40	intergenic intronic/n				
5	2450732	C	T	59	0.87	38	oncoding	AT5G07700			
5	2454751	C	T	64	0.88	40	CDS	AT5G07710	Syn	L	L
5	2486468	C	T	26	0.93	40	intergenic intronic/n				
5	2508220	C	T	44	0.88	40	oncoding	AT5G07842			
5	2531039	C	T	62	0.83	40	CDS	AT5G07930	Nonsyn	P	S
5	2646127	C	T	53	0.95	40	intronic/n oncoding	AT5G08230			
5	2785739	C	T	64	0.91	40	CDS	AT5G08590	Nonsyn	S	L
5	3077886	C	T	59	0.86	40	CDS	AT5G09870	Nonsyn	S	F
5	3269438	C	T	64	0.91	40	CDS	AT5G10390	Nonsyn	G	E
5	3297090	C	T	45	0.83	40	CDS	AT5G10470	Syn	P	P
5	3422067	C	T	44	0.86	40	CDS	AT5G10820	Nonsyn	V	M
5	3477286	C	T	48	0.84	40	CDS	AT5G10990	Nonsyn	R	W
5	3606362	C	T	22	0.96	40	intergenic				

5	4017212	C	T	57	0.9	40	CDS	AT5G12400	Nonsyn	T	I
5	4025076	C	T	44	0.85	40	CDS	AT5G12420	Nonsyn	A	T
5	4056217	C	T	38	0.84	40	intronic/n oncoding	AT5G12850			
5	4136159	C	T	55	0.87	40	intronic/n oncoding	AT5G13030			
5	4141967	C	T	10	0.71	40	intergenic				
5	4358564	C	T	42	0.82	40	CDS	AT5G13550	Nonsyn	G	E
5	4422944	C	T	47	0.87	40	CDS	AT5G13700	Nonsyn	A	T
5	4701808	C	T	39	0.93	40	intronic/n oncoding	AT5G14580			
5	4751336	C	T	45	0.8	40	CDS	AT5G14720	Nonsyn	G	E
5	5069593	C	T	48	0.83	40	CDS	AT5G15580	Nonsyn	G	R
5	5092346	C	T	60	0.91	40	CDS	AT5G15650	Syn	I	I
5	5144009	C	T	32	0.89	40	3prime_U TR	AT5G15760			
5	5243092	C	T	67	0.92	40	CDS	AT5G16040	Nonsyn	V	I

(1) Chr: chromosome. (2) Position: position of the mutated nucleotide. (3) R: nucleotide in the reference genome (pGAS1::FT ft-10 tsf-1). (4) M: nucleotide in lgf58. (5) N: number of reads supporting the mutation. (6) AF: allele frequency. (7) Sh: SHORE Score (max. 40). (8) Region: region of the locus where the mutation was identified. (9) Gen ID: gene identifier. (10) Type: type of mutation (nonsynonymous or synonymous). (11) AR: amino acid in the reference genome (pGAS1::FT ft-10 tsf-1). (12) AM: amino acid in lgf58.

1 **Supplemental Table II. List of primers used in this work**

RT-qPCR

<u>Target⁽¹⁾</u>	<u>Fw/Rv⁽²⁾</u>	<u>Sequence⁽³⁾</u>
AP1	Y28	ATGAGAGGTACTCTTACGCCGA
	Y29	CAAGTCTTCCCCAAGATAATGC
LFY	K228	TGAACATCGCTTGTCTCAT
	K229	CGACGATCCGGTACAGCTA
SOC1	K288	GTGATCTCCACTCAACAAAAA
	K289	CAACAAGAGAGAAGCAGCTTTA
PNY	K481	CAACAACCCATCTTCGTCCT
	K482	CCTCCGTTTGTGCTGCTATT
SPL4	K221	CATCATTCAAGCGACCACAG
	K222	TTGGCAAGGAAAAGCTAGGA
BOP1	K574	CGACATCCTTCGAACCCTAA
	K575	GCTCGTGTGTTTCGTCTTTCA
BOP2	K576	GGAAGGTATGAGTCGGCATC
	K577	TGCATGCCCCTCTTCTTAAT
FD	MR13	CATCAACCTTGCTTCCATCC
	MR14	GGTTTTGGTTGTGGTGGTTT
PEX4	K007	TTACGAAGGCGGTGTTTTTC
	K008	GGCGAGGCGTGTATACATT

pny-58 genotyping

<u>Target⁽¹⁾</u>	<u>Fw/Rv⁽²⁾</u>	<u>Sequence⁽³⁾</u>
PNY	K617	TCCATGTGACGTTTTGAAGG
	K618	TTCGAATGATCCCATCACAG

ChIP-qPCR

<u>Target⁽¹⁾</u>	<u>Fw/Rv⁽²⁾</u>	<u>Sequence⁽³⁾</u>
SOC1 (+)	K523	TGATTGGCACGATTCTGAAA
	K524	GAGGTGAGGATTAATAATGATGTTG
SOC1 (-)	K288	GTGATCTCCACTCAACAAAAA
	K289	CAACAAGAGAGAAGCAGCTTTA
BOP1 (1)	X272	AAGCACTTCTTCTGTCTCAT
	X273	AGAAAAGCTGGAGTTTCCAG
BOP1 (2)	X274	TCATGTCGGTAAGACGTGT
	X275	TCCTAGGGTTTTGCTTTCTG
BOP1 (3)	X264	GAGAGAGAGTGAAAAGACAA
	X265	CCCAAATCCATCAATTTTTG
BOP1 (4)	X268	GCGCCACTAATAACTTTATGG
	X269	CGTTAATTAAGTTCAGGAGC

BOP1 (5)	X270	GTCATTTCTCTCTAAACTCT
	X271	AGATCTGAATGGGCGGACCG
BOP2 (6)	X282	GAGAAAAGTAGTCTCCAAACTCTCG
	X283	ATCGTCGTGATTGGCCTAGT
BOP2 (7)	X284	ACGACTGTCAGTGCCCTTCT
	X285	TTATCTACGTCGTGCGTTCG
BOP2 (8)	X290	GTGTTGCTCAGGCTTTCACA
	X291	GAATACAAAGGTGGGCCAAA
BOP2 (9)	X292	CGAAACGCTTTTGATTTCGAT
	X293	TCGTCTGCTTTCGGAAACTT
LFY	X294	TGCATGCATTACACATAGTACACAT
	X295	TATTATCCGCCGAGCAATAGACTGTA
TOE1	X296	CCATCATGGTAAGTGGTAACCAAGTC
	X297	GAGACCCATTATTGGGAGTAACCAAA

In situ probes

<u>Target⁽¹⁾</u>	<u>Fw/Rv⁽²⁾</u>	<u>Sequence⁽³⁾</u>
PNY	K481	CAACAACCCATCTTCGTCCT
	K483	TAATACGACTCACTATAGGGCCTCCGTTTGTGCTGCTATT
BOP2	K576	GGAAGGTATGAGTCGGCATC
	K640	TAATACGACTCACTATAGGGTGCATGCCCCCTCTTCTTAAT
FD	FDT3-2F	ATTAACCCTCACTAAAGGGATTTTCATCCTCATCACCATCG
	FDT7-2R	TAATACGACTCACTATAGGG ACCAGAGCCTCGAAAGAGGT

Molecular cloning of pPNY::Venus:PNY

<u>Target⁽¹⁾</u>	<u>Fw/Rv⁽²⁾</u>	<u>Sequence⁽³⁾</u>
PNY-cds	A1-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCTTTGATATACATATTGATC GTGCTTCAAAAAGAC
	A2-R	GGGGACCACTTTGTACAAGAAAGCTGGGTCTTATTGAATCCAATTTTCATT TCTTAAAAAGATAACATTTG
PNY-prom	A3-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGGCCCTAAATATTTTG TTTTTAAAAAAAATGA
	A4-R	GGGGACCACTTTGTACAAGAAAGCTGGGTCTAGAGGAGGAGGGTGAAA GTGGAAGT
I-PIPE-1	A5-F	CATCTCATCGACATCATCCTTTCCCATGGTGAGCAAGGGCGAGG
	A6-R	CGCTGCCGCAGCGGCAGCAGCCGCAGCGCCCTTGTACAGCTCGTCCATGC CG
I-PIPE-2	A7-F	TGCCGCTGCGGCAGCGGCTGATGCATACGAGCCTTATCATGTT
	A8-R	TCTTATAATGCCCACTTTGTACAAGAAAGCTGGGTCTTATTGAATCCAATT TCATTTCTTAAAAAGATAA
V-PIPE	A9-F	TTATCTTTTAAAGAAATGAAATTGGATTCAATAGGACCCAGCTTTCTTGTA CAAAGTGGGCATTATAAGA
	A10-R	CCTCGCCCTTGCTCACCATGGGAAAGGATGATGTCGATGAGATG

- 3 (1) Target: gene, genomic region or DNA fragment flanked by the given primers. (2) Fw/Rv: forward and reverse
4 primers. (3) Sequence: DNA sequence 5' > 3' direction.